



**Euphresco**  
Phytosanitary ERA-NET



## WP5

# Asymptomatic infections of *E. amylovora* and environmental fitness

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## WP5 Problem

- *E. amylovora* exhibits different **survival strategies** to face stress environmental or plant conditions
- Many aspects of its life cycle remain unclear
- Its ability to survive in plants and to produce **latent infections** can lead to false negative results
- It is essential for giving scientific support to phytosanitary decisions:
  - the determination of survival mechanisms displayed by this pathogen under different situations
  - their involvement in fire blight epidemiology
  - the optimization of protocols for *E. amylovora* detection in the environment and asymptomatic plant material



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## WP5 Objective

**Evaluate the presence and the physiological status of *E. amylovora* in asymptomatic plant material or under stress conditions**

- Optimization of detection methods
- Implications in fire blight epidemiology and phytosanitary decision-making



## WP5 Approach

### 1. Perform **surveys to determine epidemiological significance of asymptomatic infection** and/or survival strategies of *E. amylovora*

- Harvest different sets of plant material from naturally and severely affected trees by fire blight (as a control) and asymptomatic plant material from close asymptomatic trees
- Analyze them by cultural and molecular methods

### 2. Assess **differential expression of *E. amylovora* cells under stress conditions** by high-throughput RNA-seq technologies to determine if survival strategies adopted by the pathogen have a genetic base

This part is under work



## 1a. EPPO SURVEYS TO DETECT *E. amylovora* IN ASYMPTOMATIC PLANTS

### Survey on procedures implemented in labs for analysis of asymptomatic plants for detection and identification of *E. amylovora*

- Panel on Diagnostics in Bacteriology (Sofia, 2011): survey organized on the sampling and testing procedures for asymptomatic material.
- A questionnaire developed by IVIA, ES and posted online on April 2012.

#### Sampling procedure for testing PM 7/20 (2004)

- 6 laboratories (Belgium, the Netherlands, Portugal, Slovakia, Spain and United Kingdom) perform both single plant analyses and composite samples.
- 4 laboratories (Austria, Bulgaria, the Czech Republic and Switzerland) perform single plant analyses only.
- 3 laboratories (France, Lithuania and Slovenia) perform composite sample analysis only.

Nb of bulked units	3	4	5	10	30	100
Country of the respondent	Belgium	Portugal	Spain	France	United Kingdom	Lithuania, The Netherlands, Slovakia, Slovenia



## Techniques used for testing asymptomatic samples

TECHNIQUE	Nº LABS	REFERENCE
Isolation	8	<i>Austria, France, the Netherlands, Portugal, Spain, United Kingdom, Bulgaria, Czech Republic</i>
Enrichment isolation	9	<i>Austria, Portugal, Slovenia, Spain, Czech Republic, Belgium, Lithuania, The Netherlands, Slovakia</i>
Immunofluorescence	7	<i>Czech Republic, France, United Kingdom, Bulgaria, Lithuania, The Netherlands, Slovakia</i>
Enrichment-DASI- ELISA	1	<i>Spain</i>
PCR	7	<i>Czech Republic, Spain, Austria, Belgium, The Netherlands, Portugal, Slovakia</i>
Enrichment-PCR	4	<i>Portugal, Lithuania, Slovakia, Spain</i>
Real-time PCR	6	<i>Austria, Lithuania, Slovakia, Spain</i>
Bioassay	7 +(1[2])	<i>Austria, Belgium, Portugal, Bulgaria, Czech Republic, Lithuania, The Netherlands, Slovakia</i>
Other	3	<i>Belgium, Czech Republic, The Netherlands</i>

### Number of positive *E. amylovora* tests/ Number of tests performed per year

Country of the respondent	2007	2008	2009	2010	2011
Austria	2095/3345	2369/4495	2567/3918	2145/5839	1220/4936
Belgium	0/0	0/0	3/97	0/139	0/184
Bulgaria	73/491	6/360	2/569	1/349	0/224
Czech Republic	4/31	7/53	12/68	13/155	17/141
France	0/62	0/50	0/179	0/211	0/220
Lithuania	54/334	3/273	1/173	0/130	0/2
Netherlands	1/640	1/680	2/652	12/646	6/660
Portugal	0/200	0/200	0/0	11/26	7/112
Slovakia	18/203	7/162	1/45	10/68	3/99
Slovenia	1/27	1/30	0/30	0/34	0/52
Spain	0/1376	0/2385	0/5115	4/2529	39/6164
Switzerland	1/1	1/1	1/1	1/1	1/1
United Kingdom	3/83	21/172	0/53	1/38	5/113

## *E. amylovora* positive samples

Laboratory	Techniques utilised for screening	Type of sample analysed	Composition of sample	N° of plants	Protocol used for extraction from plant material
Austria	PCR	blossom, shoots, buds, rootstocks, canker, bees	single		PM 7/20
Belgium	Isolation	buds	single & composite	3	200 buds are taken on each sample (100 twigs per sample or type plant).
Bulgaria	PM 7/20	blossom, shoots, buds, twigs leaves, green fruits, trunk	single		PM 7/20
Czech Republic	Isolation	shoots	single		PM 7/20
Lithuania	IF and PCR	blossom, shoots, twigs	single & composite	30	According to PM 7/20, appendix II.
Netherlands	Isolation	shoots	composite	100	modified version of PM 7/20
Portugal	Isolation, Nested PCR	blossom, shoots, buds	single & composite	4	PM 7/20
Slovakia	IF, PCR, Enrichment PCR	blossom, shoots, buds	single & composite	100	A. Extraction from blossom and buds (or single shoots samples). B. Extraction from shoots (composite samples of 100 plants).
Slovenia	Enrichment isolation, Enrichment-real time PCR	blossom, shoots	composite	100	EPPO 1992 and 2004 with minor modifications
Spain	PM 7/20	shoots	single & composite	5	PM 7/20,2004
Switzerland	Plating on KB/NSA, Immunoassay	blossom, shoots, buds, woody tissue, cankers	single		
United Kingdom	Isolation and IF	Twigs	single & composite	30	PM 7/20(1)

**In several EPPO countries, *E. amylovora* has been detected in asymptomatic plants using different techniques**



# 1 b. Protocol for evaluation of different numbers of samples for the analysis of asymptomatic plants for *E. amylovora*

## Preparation and analyses of SPIKED SAMPLES

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4 g healthy plant material in 200 ml of antioxidant buffer (EPPO 2013)



Add bacterial suspensions of *E. amylovora* strain to 25 ml extracts  
(final concentrations: 10, 10<sup>2</sup>, 10<sup>3</sup>, 10<sup>4</sup>, 10<sup>5</sup> and 10<sup>6</sup> cfu/ml)



Isolation and techniques of EPPO protocol  
Keep 5 ml at - 20°C, and add 30% glycerol to the remaining amount for bulk samples



Enrichment in King's B and CCT media



Isolation on CCT, ELISA, Real time-PCR or Conventional PCR



Calculate the sensitivity of the protocol in each country  
(Individual samples)





## Preparation and analyses of BULK SAMPLES

0.1 g of the same type of plant material from each 2, 4, 9 or 19 healthy plants of the same host (samples b1, b2, b3 or b4)



Add 6.5ml of each of the positive defrosted samples



Enrichment in King's B and CCT media



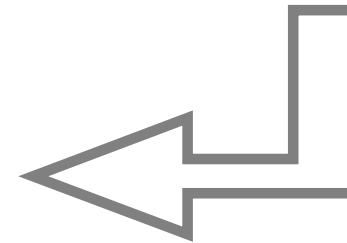
Isolation on CCT, ELISA, Real time-PCR or Conventional PCR



Calculate the sensitivity of the protocol with the 3, 5, 10 or 20 samples of plant material

## SPIKED SAMPLES

Defrost samples that gave positive result



# Results

## Spiked samples

### IVIA – Assay 1 Pear Conference



		Neg	10 <sup>2</sup>	10 <sup>3</sup>	10 <sup>4</sup>	10 <sup>5</sup>	10 <sup>6</sup>	10 <sup>7</sup>	Ea control
Isolation		-	+	+	+	+	+	+	+
E-ELISA-DASI	KB	-	-	-	-	-	-	-	+
	CCT	-	+	+	+	+	+	+	+
rt-PCR (Pirc et al, 2009)		-	-	-	+	+	+	+	+

**Sensitivity: 10<sup>2</sup> cfu/ml for isolation and ELISA DASI (after enrichment in CCT)**  
**10<sup>4</sup> cfu/ml for real-time PCR**

## Bulk samples

b1: sample consisting in 3 plants ; b2: 5 plants; b3: 10 plants; b4: 20 plants

		10 <sup>2</sup>				10 <sup>3</sup>				10 <sup>4</sup>			
		b1	b2	b3	b4	b1	b2	b3	b4	b1	b2	b3	b4
Isolation	KB	+	+	+	ND	+	+	+	ND	+	+	+	ND
	CCT	+	+	+	-	+	+	+	+	+	+	+	+
E-ELISA-DASI	KB	+	-	+	ND	+	+	+	ND	+	+	+	ND
	CCT	+	+	+	-	+	+	+	+	+	+	+	+
rt-PCR (Pirc et al, 2009)	KB	+	+	+	ND	+	+	+	ND	+	+	+	ND
	CCT	+	+	+	+	+	+	+	+	+	+	+	+

**Sensitivity: 10<sup>2</sup> cfu/ml in a sample consisting on 1, 3, 5 or 10 plants for all techniques(after enrichment)**

# Results

## IVIA – Assay 2 Pear Blanquilla



### Spiked samples

		Neg	10 <sup>1</sup>	10 <sup>2</sup>	10 <sup>3</sup>	10 <sup>4</sup>	10 <sup>5</sup>	10 <sup>6</sup>	10 <sup>7</sup>	Ea control
Isolation		-	+	+	+	+	+	+	+	ND
E-ELISA-DASI	KB	-	+	+	-	+	+	+	+	ND
	CCT	-	-	-	+	+	+	+	+	ND
rt-PCR (Pirc et al, 2009)		-	-	-	+	+	+	+	+	ND

**Sensitivity: 10 cfu/ml for isolation and ELISA DASI (enrichment in KB)**  
**10<sup>3</sup> cfu/ml for real-time PCR**

### Bulk samples

b1: sample consisting in 3 plants ; b2: 5 plants; b3: 10 plants; b4: 20 plants

		10 <sup>1</sup>				10 <sup>2</sup>				10 <sup>3</sup>			
		b1	b2	b3	b4	b1	b2	b3	b4	b1	b2	b3	b4
Isolation	KB	+	+	+		+	+	+		+	+	+	
	CCT	+	+	+		+	+	+		+	+	+	
E-ELISA DASI	KB	-	-	+		+	+	+		+	+	+	
	CCT	+	+	-		+	+	+		+	+	+	
rt-PCR (Pirc et al, 2009)	KB	+	+	+		+	+	+		+	+	+	
	CCT	+	+	-		+	+	+		+	+	+	

**Sensitivity: 10 cfu/ml in a sample consisting on 1, 3, 5 or 10 plants for isolation, ELISA, DASI and rt-PCR (after enrichment in KB)**

## 1.c. Analysis of asymptomatic loquat samples from a Spanish area where *E. amylovora* was detected

Cooperative work with Plant Protection Service of Comunidad Valenciana (Spain) and Cooperative Ruchey, callosa d'Ensarià, Spain

### Materials and Methods: OEPP Protocol

### Results

	November '13	January '14	March '14	
Nº analysed samples	Plot 1- 142 Blossom Plot 2- 122 Blossom	Plot 1- 61 Blossom 61 Fruit 61 Shoot	Plot 3- 30 Fruit-shoot Plot 4- 39 Fruit-shoot	516
Nº E-ELISA +	3 3	0 0 0	0 0	6
Nº real time PCR +	21 4	ND ND ND	2 (1 fruit-1 shoot) 0	21
Nº Taylor PCR +	6 1	ND ND	ND ND	7
Isolation	0	0	0	0

**Negative isolation in winter, but positive detection by ELISA, conventional and real-time PCR in November. Probably latent infections.**



## WP5 Deliverables

- Optimized protocol for detection of *E. amylovora* in asymptomatic plant material, based on the results obtained in the different countries.
- Survival ability of *E. amylovora* under adverse conditions that can promote non-detection by conventional methods evaluated.
- Genes involved in the response of *E. amylovora* against determined adverse conditions.



## WP5 Dissemination and training output

- Protocol with advices and the most suitable methods for detection of *E. amylovora* in asymptomatic plant material.
- A transcriptome of *E. amylovora* under copper stress:
  - \*Águila-Clares, B., Marco-Noales, E., López, M.M., Sundin, G.W. 2013. How does *Erwinia amylovora* face up to stress by copper? 13th International Workshop in Fire Blight. Zurich.
  - \*Águila-Clares, B., Marco-Noales, E., López, M.M., Sundin, G.W. 2013. Análisis transcriptómico de la bacteria *Erwinia amylovora* en respuesta al estrés por cobre. XXIV Congreso de Microbiología SEM. L'Hospitalet, Barcelona.
- Scientific publication in a peer-reviewed journal describing the survival ability of *E. amylovora* in asymptomatic plant material and an optimized protocol for its detection.
- Scientific publication in a peer-reviewed journal on the genetic basis of survival response of *E. amylovora* to certain stress conditions: Águila-Clares B, Marco-Noales E, Penyalver R, López MM, Sundin GW. Genetic systems operating in *Erwinia amylovora* after a copper shock: the role of *copA*. Manuscript in preparation for Molecular Plant Pathology.
- Article in a popular science magazine translating research in an accessible format for e.g., plant health inspectors, diagnostic laboratories, pome fruit growers, and extension workers: Roselló M, Gamón M, Ferrer A, Dalmau V, Palacio-Bielsa A, López MM. 2014. Prevención del fuego bacteriano en plantaciones de níspero (*Eriobotrya japonica*) en la Comunitat Valenciana. Phytoma 259: 1- 5.



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**Thanks for your attention!!**